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1 **Synchronous genetic turnovers across Western Eurasia in Late Pleistocene**
2 **collared lemmings**

3 Running head: biogeographic history of the collared lemming

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54 **Abstract**

55 Recent palaeogenetic studies indicate a highly dynamic history in collared lemmings
56 (*Dicrostonyx* spp.), with several demographic changes linked to climatic fluctuations that took
57 place during the last glaciation. At the western range margin of *D. torquatus*, these
58 demographic changes were characterized by a series of local extinctions and recolonizations.
59 However, it is unclear whether this pattern represents a local phenomenon, possibly driven by
60 ecological edge effects, or whether these extinctions and recolonizations took place across
61 large geographical scales. To address this, we explored the palaeogenetic history of the
62 collared lemming using a next-generation sequencing approach for pooled mitochondrial
63 DNA amplicons. Sequences were obtained from over 300 fossil remains sampled across
64 Eurasia and two sites in North America. We identified five mitochondrial lineages of *D.*
65 *torquatus* that succeeded each other through time across Europe and western Russia,
66 indicating a history of repeated population extinctions and recolonizations, most likely from
67 eastern Russia, during the last 50,000 years. The observation of repeated extinctions across
68 such a vast geographical range indicates large-scale changes in the steppe-tundra environment
69 in western Eurasia during the last glaciation. All Holocene samples, from across the species'
70 entire range, belonged to only one of the five mitochondrial lineages. Thus, extant *D.*
71 *torquatus* populations only harbour a small fraction of the genetic diversity that existed during
72 the Late Pleistocene. In North American samples, haplotypes belonging to both *D.*
73 *groenlandicus* and *D. richardsoni* were recovered from a Late Pleistocene site in
74 southwestern Canada. This suggests that *D. groenlandicus* had a more southern, and *D.*
75 *richardsoni* a more northern glacial distribution than previously thought.

76 **Introduction**

77 The Late Pleistocene was a characterized by dramatic shifts in mammal species' ranges
78 (Hewitt, 1999), changes in community composition and richness (Blois *et al.*, 2010, Graham,
79 1986, Stewart, 2009), loss of genetic diversity (Shapiro *et al.*, 2004, Stiller *et al.*, 2010),
80 population replacements (Hofreiter *et al.*, 2007, Leonard *et al.*, 2007) and local or global
81 extinctions (Martin, 1984, Stiller *et al.*, 2014). Individual taxa have responded differently to
82 the effects of climatic and environmental changes (Lorenzen *et al.*, 2011, Prost *et al.*, 2013,
83 Stewart *et al.*, 2010). Different responses have also been observed within the same species at
84 the population level (Campos *et al.*, 2010). While many studies have focused on a single or a
85 restricted number of populations to explore their dynamics and identify the forcing
86 mechanisms, the identified trends are not necessarily characteristic of the entire species.
87 Specific factors, such as local environmental changes can play an important role in shaping
88 population dynamics. Further, different responses can be expected for core versus peripheral
89 populations due to source-sink dynamics and ecological edge effects. These factors may affect
90 the level of genetic variation and evolutionary potential of populations at the edge of species'
91 geographical ranges, depending on the amount of gene flow from the core population (Brown,
92 1984, Hampe & Petit, 2005). Species-level investigations require sampling of multiple
93 populations from across the geographic distribution in order to gain a complete understanding
94 of the evolutionary processes that led to current patterns of biodiversity.

95 Collared lemmings (*Dicrostonyx* spp.) are cold-adapted, small rodents restricted to the dry,
96 treeless environment of the Arctic tundra (Kowalski, 1995). Their present-day distribution is
97 nearly circumpolar with *D. torquatus* occupying a range from western Russia to northeast
98 Siberia, *D. groenlandicus* inhabiting Alaska, the Canadian Arctic and Wrangel Island, and *D.*
99 *richardsoni* and *D. hudsonius* occupying regions west and east of Hudson Bay respectively.

100 The recognition of one Eurasian species and three North American species is based on

101 mitochondrial DNA (mtDNA) diversity, karyotype variation and hybridization experiments,
102 and represents the most widely accepted taxonomic classification (Fedorov & Goropashnaya,
103 1999). Evidence of a wider geographical range both in Eurasia and North America during the
104 Late Pleistocene has been provided by the fossil record; collared lemmings appear to have
105 expanded southwards during cold periods while they remained restricted to the north during
106 warm interglacial periods, including the present one (Graham *et al.*, 1996, Sher, 1991,
107 Stewart, 2003). It has also been shown that the climates associated with the expanded
108 populations, living in non-analogue communities, are not necessarily equivalent to those
109 associated with modern populations to the North. This may imply that these southern
110 populations were differently adapted to modern ones (Stewart, 2009, Stewart *et al.*, 2003).
111 Historical population fragmentation associated with the glacial/interglacial cycles has been
112 inferred from the observed phylogeographic structure reconstructed from analysis of modern
113 mtDNA data (Fedorov *et al.*, 1999, Fedorov, 1999). Moreover, the observed low nucleotide
114 and haplotype diversity in the mtDNA of *D. torquatus* has been attributed to regional
115 bottlenecks most likely linked to warming events during the Holocene (Fedorov *et al.*, 1999).

116 Late Pleistocene vertebrate sites are rich in small mammal remains (Markova *et al.*, 1995,
117 Stewart *et al.*, 2003), and these constitute a valuable source of ancient DNA that enables
118 reconstruction of past population histories. Previous genetic studies have exploited the
119 abundance of collared lemming fossil remains collected from regions within the genus'
120 current and past distribution to identify distinct demographic histories in populations from
121 different sites. For instance, genetic continuity during the last 25,000 years has been reported
122 for *D. torquatus* from a single site in the northern Urals (Pymva Shor) with signatures of a
123 severe population bottleneck following the Last Glacial Maximum (LGM) most likely due to
124 climate warming (Prost *et al.*, 2010). Conversely, a series of local population extinctions
125 followed by recolonization events has been documented at the westernmost range of *D.*

126 *torquatus* during the last 50,000 years (Brace *et al.*, 2012). These repeated turnover events
127 were correlated with periods of climatic fluctuation during the last glaciation, and thus likely
128 reflected response to environmental changes. Although both studies demonstrated that climate
129 changes played a major role in driving the dynamics of *D. torquatus*, they indicated the
130 possibility of distinct patterns of response in different parts of the species' distribution, with a
131 single demographic bottleneck in the center and several local extinctions/replacements near
132 the edge of its range.

133 In North America, species-level identification of fossils of *Dicrostonyx* is hampered by
134 similarities in molar morphology among extant forms, except for *D. hudsonius*, which
135 displays diagnostic morphological characters (Fulton *et al.*, 2013). Evaluation of ancient DNA
136 provides an alternative method for distinguishing morphologically similar taxa but thus far
137 very few North American fossil remains of *Dicrostonyx* have been genetically studied.
138 However, a study on mtDNA from three individuals from Midwest United States dated to
139 ~15,000 and ~24,000 years ago (Fulton *et al.*, 2013) revealed that they belong to *D.*
140 *richardsoni*. This finding led the authors to hypothesize glacial survival of the species at
141 regions south of the Laurentide ice sheet, similar to what has been suggested for *D. hudsonius*
142 (Macpherson, 1965). However, it remains unknown whether *D. groenlandicus* also was
143 restricted to regions south of the Laurentide ice sheet during the last glacial period or if this
144 type of geographic response was specific to *D. richardsoni* and *D. hudsonius*.

145 In this study, we broadened the genetic sampling of fossil remains of *Dicrostonyx* to cover
146 most of its Eurasian Late Pleistocene range. We further analysed ancient specimens from two
147 cave sites in Alberta in the Canadian Rocky Mountains, a region that has remained
148 unexplored in terms of small mammal palaeogenetic analyses. Using ancient DNA methods,
149 we generated a dataset of mitochondrial sequences from more than 300 individuals. We
150 combined our dataset with previously published modern and ancient data to address several

151 questions regarding the past population dynamics and biogeography in collared lemmings.
152 First of all, we evaluated the possibility that local extinctions and recolonizations were
153 widespread across the range of *D. torquatus* during the Late Pleistocene. Moreover, we were
154 interested in exploring whether any observed extinctions and recolonizations were
155 synchronous in time across several geographic sites, since this may indicate extensive changes
156 in the glacial steppe-tundra environment. Finally, we used our data to determine which
157 species of collared lemming occupied southwestern Canada during the last glacial period,
158 thereby providing additional data to assess Pleistocene origins of extant North American
159 populations of *Dicrostonyx*.

160

161 **Materials and methods**

162 *Sample collection and DNA analyses*

163 We collected 621 collared lemming mandibles and molars from across Eurasia and western
164 Canada covering a temporal range from present day to more than 50,000 calendar years
165 before present (cal. years BP; Supporting Table S1). To facilitate discussion of the Eurasian
166 dataset we refer to localities in Russia west of the Ural Mountains as western Russia,
167 localities in north-central Siberia as Central Russia and localities in northeastern Siberia as
168 eastern Russia. DNA extractions were conducted in two laboratories dedicated for ancient
169 DNA, housed at the Swedish Museum of Natural History and the Institute of Genetics and
170 Biotechnology at the University of Warsaw. Amplification and sequencing of the
171 mitochondrial cytochrome *b* (cyt *b*) gene was performed with barcoded primers or library
172 adapters to enable parallel sequencing of pooled individuals using the 454 sequencing
173 technology. For full details on methods and data analyses, see Supporting Information.

174 *Radiocarbon dating*

175 We radiocarbon dated specimens with enough remaining material (more than 50mg after
176 material had been taken for DNA extractions) and for which the entire sequence of targeted
177 mtDNA was obtained. Dating was performed with the Accelerator Mass Spectrometry (AMS)
178 method at the Oxford Radiocarbon Accelerator Unit using methods previously outlined
179 (Brock *et al.*, 2010) (Supporting Table S2). OxCal v. 4.2 (Ramsey, 2009) and the IntCal 13
180 calibration curve (Reimer *et al.*, 2013) were used to calibrate the radiocarbon dates obtained
181 in this study as well as previously published dates. Calibrated median dates are given in
182 Supporting Tables S2, S3.

183 *Phylogenetic analyses*

184 We aligned sequences generated in this study with previously published ancient and modern
185 complete *cyt b* sequences (see Supporting Table S3, for GenBank accession numbers) in
186 Geneious version 5.5.7. The best-fitting model of nucleotide substitution for each dataset was
187 chosen by MrModetest2 (Nylander, 2004) using a partition scheme with three coding
188 positions. We performed three sets of analyses: *i*) using a dataset comprised of single
189 haplotypes (identical sequences were collapsed), *ii*) using a dataset restricted to dated
190 sequences only (with finite radiocarbon dates), excluding North American sequences and *iii*)
191 using a dataset including radiocarbon dated sequences as well as sequences for which prior
192 information on their age was available. Bayesian phylogenies were constructed in BEAST
193 v.1.8.0 (Drummond *et al.*, 2012) implementing a strict molecular clock with the substitution
194 rate estimated from the data. Analysis of dataset (*i*) was performed under the speciation: birth-
195 death tree prior given that more than one species were included. For dataset (*ii*), we compared
196 three population models, namely constant population size, Bayesian skyline plot (Drummond
197 *et al.*, 2005) and Bayesian SkyGrid (Gill *et al.*, 2013) using marginal likelihood estimation
198 (MLE) (Baele *et al.*, 2013). The Bayesian SkyGrid model was the model of choice from the
199 MLE method and implemented in the final analyses. In datasets (*ii*) and (*iii*) we used the
200 calibrated median of the radiocarbon age of each sequence as tip-dates for internal calibration
201 of the tree, even in cases where the calibrated date extended out of the IntCal13 curve range.
202 Sequences that provided infinite radiocarbon dates were excluded from these datasets. In
203 dataset (*iii*), undated sequences as well as sequences for which radiocarbon dating failed were
204 given a uniform distribution of dates as a prior for age sampling, based on stratigraphic
205 information or associated dated material (field 'Prior age' in Supporting Table S2). Sequences
206 for which prior information on their age was not available were excluded from this dataset. To
207 test whether the temporal span of the dated samples and the sequence information content
208 were sufficient to calibrate the tree and estimate evolutionary rates, we performed date

209 randomization test as described (Ho *et al.*, 2011) (Fig. S1). The GTR+I+G model of sequence
210 evolution was chosen by MrModeltest2 for datasets (i) and (iii) using both hierarchical
211 likelihood ratio tests (hLRTs) and the Akaike Information Criterion (AIC). However, due to
212 poor estimates of convergence even after increasing the number of generations to 100 million,
213 the implemented model was changed to HKY+I+G. For dataset (ii) we implemented the
214 HKY+I+G model of sequence evolution, which was the best-fitting model according to the
215 AIC (hLRTs recommended HKY+G). Two independent runs of 100 million generations (50
216 million for dataset [ii]) were performed for each analysis sampling every 10,000 generations
217 (5,000 for dataset [ii]) and discarding the first 10% of the samples as burnin. LogCombiner
218 v.1.8.0 was used to combine log files and tree files, and Tracer v.1.6. (Rambaut *et al.*, 2014)
219 was used to assess convergence between runs.

220 We also constructed a Bayesian SkyGrid Plot (BSG) (Gill *et al.*, 2013) in BEAST using
221 dataset (ii) to infer changes in female effective population size (N_{ef}) through time. Two
222 independent runs of 50 million iterations were run with sampling every 5,000 generations and
223 discarding the first 10% of the samples as burnin. Tracer v.1.6. was used to assess
224 convergence and estimate the BSG.

225 We reconstructed the spatiotemporal distribution of the mitochondrial lineages in GenGIS
226 (Parks *et al.*, 2009) by assigning dated individuals or individuals for which prior age
227 information was available, to 5 time slices divided according to the approximate temporal
228 distribution of each mitochondrial lineage.

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229 **Results**

230 *Mitochondrial DNA diversity*

231 When we screened our material for a fragment of 171bp of the targeted mtDNA region, we
232 found that DNA preservation levels varied among different locations. Certain sites exhibited
233 high frequencies of DNA-yielding samples while other sites displayed lower rates. In total
234 452 specimens yielded amplifiable DNA and 330 of these provided complete *cyt b* sequences
235 (780bp) (Table S2). Only complete *cyt b* sequences were included in the subsequent analyses.
236 We detected 159 novel haplotypes (deposited in GenBank with accession numbers XXXX;
237 Supporting Table S2). When previously published modern and ancient sequences (Supporting
238 Table S3) were combined with our data, this resulted in a dataset that comprised a total of 431
239 complete *cyt b* sequences.

240 *Radiocarbon dating*

241 Due to the small size of the starting material (not more than 200mg), most of the samples
242 were not treated with ultrafiltration but only gelatinized and filtered for AMS dating. The
243 combination of small sample size and lack of ultrafiltration resulted in a large number of
244 specimens (48 out of 120 or 40%) failing to produce radiocarbon dates because of low
245 collagen yield and/or high carbon:nitrogen ratios, which indicate degradation and
246 contamination of exogenous carbon, respectively. Successfully dated specimens produced a
247 range of dates from ~12,000 radiocarbon years BP to infinite (above ~50,000 radiocarbon
248 years indicating an age beyond the limits of radiocarbon dating). Furthermore, specimens
249 from two sites, Ostrov Bolshevik in central Russia and Kyttyk peninsula in eastern Russia
250 produced very recent radiocarbon dates (less than 1,000 radiocarbon years) confirming that
251 these specimens are modern and of Late Holocene origin respectively (Supporting Table S2).

252 *Phylogenetic structure and demography*

253 Bayesian phylogenetic analyses revealed 8 major mitochondrial lineages (Fig. 1). Of these,
254 lineages EA1 to EA5 were of Eurasian origin and belonged to the clade representing *D.*
255 *torquatus*. These lineages correspond to lineages 1-5 described in Brace *et al.* (2012).
256 Lineages NA1 to NA3 correspond to the three North American collared lemming species (*D.*
257 *groenlandicus*, *D. richardsoni* and *D. hudsonius*). All of the lineages were monophyletic and
258 supported by high to moderate posterior probability values.

259 Focusing on *D. torquatus*, a temporal structure was observed within the diversity of the
260 Eurasian lineages (Fig. 1), which was present across a large geographical scale ranging from
261 western Europe to western Russia. This temporal division is more clearly observed in the
262 phylogeny with finite radiocarbon-dated sequences (Fig. 2) and is illustrated by the
263 geographical/temporal distribution of the lineages shown in Fig. 3. The two most basal
264 Eurasian lineages (EA1 & EA2) were comprised of the oldest sequences in our dataset with
265 dates greater than ~50,000 (infinite radiocarbon dates) up to ~42,300 cal. years BP. These two
266 lineages were followed by lineage EA3, which consisted of specimens that spanned a time
267 range from ~32,000 to ~22,800 cal. years BP. Lineage EA4 was, with one exception, dated to
268 a short time period between 22,200 and ~20,500 cal. years BP, whereas lineage EA5 included
269 Late Pleistocene European and western Russian sequences younger than ~20,500 cal. years
270 old together with all Late Holocene and modern sequences from across Russia. This latter
271 lineage (EA5) covers most of the species' present-day geographical distribution. Within
272 lineage EA5, we observed some evidence of a phylogeographic structure among the modern
273 haplotypes. The haplotypes from Ostrov Bolshevik in Central Russia (H112-H118) were
274 placed within the diversity of modern *D. torquatus* but formed a distinct group (Fig. 1).
275 Contrary to indications from the morphology of their molars that these specimens are relics of
276 last glacial morphs (Abramson *et al.*, 2004), their mtDNA sequences suggest a more recent
277 origin of this population.

278 The pattern of sequential phylogenetic groups through time appears to have occurred in most
279 of the studied sites. In general, the lineages in western Russia and Europe did not overlap in
280 time, although there were a few exceptions to this pattern. The oldest sequences within
281 lineage EA5 from Biśnik cave in Poland were dated to ~19,000 cal. years BP, although one
282 specimen from the same site with a younger date (16,159 cal. years BP) actually fell within
283 lineage EA4 (Fig. 2). All other sequences within EA4 were tightly grouped in time, ranging
284 from 20,383 to 22,157 cal. years BP. Furthermore, the only finite-dated sequence within
285 lineage EA2 (44,356 cal. years BP) postdated some of the Belgian sequences within lineage
286 EA1 (Fig. 2).

287 When we included additional sequences in the phylogeny by assigning prior ranges for the
288 age of undated specimens as tip-dates, based on stratigraphic information or associated dated
289 material, this provided further support for the observed temporally-structured mitochondrial
290 diversity (Supporting Fig. S2). There was again an association between the inferred age of the
291 sequences and the mitochondrial lineage they were placed within across nearly all studied
292 sites in Europe and western Russia. One single exception to this pattern was observed in
293 Pymva Shor in western Russia, where the assumed age of sequence E333 (~26,000 cal. years
294 BP) did not fit with the observed temporal range of the lineage it fell within (EA4, up to
295 ~22,200 cal. years BP; Fig. 3, Supporting Fig. S2).

296 The phylogenetic position of ancient haplotypes from eastern Russia is noteworthy.
297 Haplotypes H169, H196 were basal to lineage EA4, and haplotypes H194, H195, H96 were
298 basal to lineages EA5-EA2 (Fig. 1, Supporting Table S2). However, it should be noted that
299 support for this latter association was very low, and in the phylogeny where sequences with
300 prior information on their age were included (Supporting Fig. S2) the position of haplotype
301 H96 (represented by sequence E157 from Bison's site in northeastern Russia) changed and
302 became basal to the entire diversity of *D. torquatus* with high support value. One more

303 specimen that stood out in the phylogeny is the one represented by haplotype H119, which
304 originated from Batagay close to the Yana River in eastern Russia, and fell at the base of the
305 entire phylogeny including both Eurasian and North American mitochondrial lineages (Fig.
306 1).

307 The ancient haplotypes from North America were phylogenetically placed within the
308 mitochondrial diversity currently observed among extant collared lemmings in Alaska and
309 Canada (Fig. 1). Haplotypes H92, H93 were basal to *D. groenlandicus* whereas haplotype
310 H121 was grouped together with *D. richardsoni*, and haplotype H120 was basal to both these
311 lineages.

312 The Bayesian SkyGrid Plot revealed changes in N_{ef} through time but was not able to capture
313 the population extinction/replacements in any detail (Supporting Fig. S3). This could be
314 attributed to the temporal (albeit not spatial) structure in our data, which violates the
315 assumption of a single panmictic population in BEAST (Drummond *et al.*, 2005, Navascues
316 *et al.*, 2010).

317 Discussion

318 We identified five mitochondrial lineages in *D. torquatus* that sequentially replaced each
319 other through time across a large part of the species' distribution (Fig. 2, 3). These five
320 lineages have previously been identified in a regional ancient DNA study on collared
321 lemmings from western Europe, where the successive replacement of these lineages through
322 time was interpreted as a series of local population extinctions followed by recolonizations
323 (Brace *et al.*, 2012). Our results demonstrate that this pattern was not limited to the western
324 edge of the species' range, but extended across a large part of its geographical distribution
325 including Europe and western Russia (Fig. 3). Thus, rather than indicating an edge effect at
326 the western limits of the species' range margin, which could have been driven by small-scale
327 ecological perturbations, our results are more consistent with a scenario of several major
328 environmental changes that affected a large part of the Eurasian steppe tundra biome.

329 At least three turnover events were revealed from the reconstructed phylogeny (Fig. 2). In
330 general, the timing of these appear synchronous across different sites in Europe and western
331 Russia (Fig. 3). Replacement of lineages EA1-EA2 by lineage EA3 appears to have occurred
332 between ~42,300 and ~32,000 cal. years BP, after which the climate started to become colder
333 following the end of Greenland Interstadial (GI) 5. Interestingly, this is a time period that has
334 been marked by population replacements in other species within Europe, such as the cave bear
335 (Hofreiter *et al.*, 2007) and the woolly mammoth (Palkopoulou *et al.*, 2013). The two
336 subsequent replacements appear to have had much shorter time ranges, with the replacement
337 of lineage EA3 by EA4 occurring between ~22,800 and ~22,200 cal. years BP, and that of
338 lineage EA4 by lineage EA5 around 20,500 cal. years BP. The time intervals of both these
339 replacements fall within the LGM when tundra and steppe tundra environments spread over
340 most of Europe and Western Asia. However, the observed turnovers seem to have been
341 preceded by the short warming stage of GI 2 (Svensson *et al.*, 2008) and moreover they

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342 coincide with short term warmings in the high resolution palynological record from Lago
343 Grande di Monticchio in Italy (Allen *et al.*, 1999) (Supporting Fig. S4). This suggests that
344 short term environmental changes could have affected a large part of the Eurasian steppe
345 tundra biome during the LGM. Moreover, it is notable that the latter replacement occurred
346 within a time period (21,700 – 19,400 cal. years BP) for which a gap has been reported in the
347 fossil record of woolly mammoths from central and northwestern Europe, when the
348 Weichselian ice sheet reached its maximum extent (Nadachowski *et al.*, 2011, Stuart *et al.*,
349 2004, Ukkonen *et al.*, 2011). As for the two oldest lineages (EA1 and EA2), it is unclear
350 whether these were temporally separated, and hence whether they represent a fourth
351 extinction/recolonization event, since they included sequences that were very close to the
352 limits of radiocarbon dating (many of these specimens provided infinite dates).

353 Interestingly, we found evidence of mitochondrial lineage replacement as far east as the high-
354 Arctic site of Pymva Shor in the northern Urals. Mitochondrial DNA variation data from this
355 site was previously interpreted as indicating population continuity through time (Brace *et al.*,
356 2012, Prost *et al.*, 2010). However, our data suggests a replacement of lineage EA4 by lineage
357 EA5 sometime between ~16,000 and ~26,000 cal. years BP (Fig. 3). We hypothesize that the
358 difference between our results and those from previous studies can be explained by higher
359 accuracy offered by the comparatively longer *cyt b* sequences retrieved in our study. Prost *et*
360 *al.* (2010) used a fragment of similar size (704bp) that included 282bp *cyt b* and 426bp control
361 region (CR). However, CR shows less diversity in collared lemmings than *cyt b* (Prost *et al.*,
362 2010) and thus a longer fragment of the *cyt b* should result in an increase in resolution.
363 Overall, the finding that lineage replacements were geographically widespread indicates a
364 pattern of repeated population extinctions across much of the collared lemming's Eurasian
365 distribution.

366 However, there were a few exceptions to the overall pattern of synchronous replacements, and
367 these merit a closer discussion. In Biśnik cave in Poland, the presence of an ~16,000 years old
368 sequence (95% range: 15,901-16,411 cal. years BP) within lineage EA4 (Fig. 2, Fig. 3) could
369 indicate that lineage EA4 coexisted with lineage EA5 (for at least 4,000 years taking into
370 account the error ranges of the calibrated dates). However, this is only a single sequence and
371 more data are needed to resolve whether these two lineages overlapped in time in this region.
372 Moreover, the phylogenetic placement and associated date of the oldest sequence from Pymva
373 Shor in western Russia indicates a larger upper temporal range for lineage EA4, which could
374 further imply that lineages EA4 and EA3 temporally overlapped in this region (Fig. 3,
375 Supporting Fig S2). However, the lack of older specimens from Pymva Shor does not allow
376 us to explore the mitochondrial diversity that existed further back in time. Alternatively,
377 temporal discrepancy in Pymva Shor could be due to errors in the assumed age of the
378 specimens, since these were based on associated rather than direct radiocarbon dates
379 (Golovachov & Smirnov, 2009). Sequencing of additional directly-dated specimens from
380 Pymva Shor is needed to unravel the timing of the replacement of lineages EA4 and EA3 at
381 that site. Nonetheless, the phylogenetic placement of specimens from other western Russian
382 sites (e.g. Yudinovo, Studenaya, Betovo), which were directly dated and altogether cover a
383 wider temporal range (Fig. 2, Fig. 3), reinforces the observed pattern of simultaneous lineage
384 turnovers in western Russia and Europe.

385 Genetic replacements, indicating population extinctions and recolonizations, have been
386 observed in several other taxa (Barnes *et al.*, 2007, Hofreiter *et al.*, 2007, Leonard *et al.*, 2007,
387 Stiller *et al.*, 2014). However, most of these studies were conducted on samples from limited
388 geographical distributions, and it is unclear whether these events were site-specific or more
389 widespread. In contrast, our study demonstrates several lineage replacements, which indicate
390 a series of extinctions and recolonisations across a vast geographical area covering Europe

391 and western Russia. It is therefore possible that population replacements occurred at a wider
392 spatial extent in other taxa too. To assess this hypothesis, further analyses on additional
393 samples from other taxa covering a broader geographical range are needed.

394 In contrast to the pattern observed in Europe and western Russia, we did not observe a similar
395 temporal separation of lineages in the eastern range of *D. torquatus* (Fig. 3, Supporting Fig.
396 S2). Although all modern eastern Russian haplotypes were phylogenetically placed together
397 with all other modern haplotypes within lineage EA5, the phylogenetic placement of Late
398 Pleistocene eastern Russian sequences do not seem to correspond to their age. Based on dates
399 from associated material, sequences L260, L261 have an inferred age between ~30,000 to
400 ~34,000 cal. years BP. Despite this, these sequences were basal to lineage EA4, where all
401 other sequences have ages ranging from ~20,500 to ~22,200 cal. years BP (Supporting Fig.
402 S2). Although our sampling across eastern Russia was limited compared to western Eurasia,
403 both in terms of sample size as well as temporal coverage, it is interesting to note that all Late
404 Pleistocene eastern Russian sequences occupy basal positions in their respective mtDNA
405 lineages. This indicates that eastern Russia was the source population from which founders
406 recolonized western Russia and Europe following each of the inferred extinctions. The
407 seemingly high genetic diversity in Late Pleistocene samples from eastern Russia is also
408 consistent with the hypothesis that Beringia served as an interglacial refugium for the cold-
409 adapted *D. torquatus* (Fedorov & Goropashnaya, 1999).

410 One of our samples, from a site near the Yana river in eastern Russia, yielded a highly
411 divergent haplotype (H119, represented by sequence E313) that was basal to the entire
412 phylogeny including both Eurasian and North American mitochondrial lineages (Fig. 1). This
413 specimen yielded an infinite radiocarbon date (>50,299; Supporting Table S2). It is tempting
414 to interpret the basal placement in the phylogeny in combination with the specimen's ancient
415 origin as representing ancestral variation that was present prior to the split between Eurasian

416 and North American collared lemmings. Alternatively, this haplotype could represent an
417 undiscovered extinct *Dicrostonyx* species. Further sequencing of this specimen including both
418 mitochondrial and nuclear DNA, as well as sequencing of additional collared lemming fossil
419 remains from eastern Russia, may help to resolve the identity of this haplotype and the role of
420 northeastern Siberia as a refugium during periods of unfavorable environmental conditions.

421 Moving towards the present, we found some evidence of contemporary phylogeographical
422 structure in *D. torquatus* (Fig. 1) in congruence with earlier findings (Fedorov *et al.*, 1999,
423 Prost *et al.*, 2010). It is noteworthy that most of the contemporary phylogeographic clades
424 across the Palearctic have also been found to exhibit variation in chromosomal numbers,
425 although the distributions of mtDNA clades and chromosomal races do not agree in absolute
426 terms (Fedorov *et al.*, 1999). These were hypothesized to have resulted from isolation during
427 previous glacial cycles while regional population reductions during the Holocene were
428 implicated for the limited mtDNA variation observed within each clade (Fedorov *et al.*,
429 1999). Our phylogenetic analyses demonstrate that the modern mtDNA clades diverged much
430 more recently than previously thought, within the last 20,000 years and that each of the clades
431 coalesced during the last 10,000 years (Fig. 2, Supporting Fig. S2). Thus, the current
432 phylogeographic pattern may be a consequence of population contraction and isolation during
433 warming periods at the end of the last glacial period. This hypothesis is consistent with
434 observed changes in tooth morphotype frequencies (Smirnov & Fedorov, 2003), which also
435 occurred during that time. Interestingly, the observation of a recent origin for the present day
436 phylogeographic groups also implies that the current chromosomal variation among extant *D.*
437 *torquatus* populations evolved recently, possibly as a consequence of Late Pleistocene
438 bottlenecks and the ensuing recolonization of the Eurasian Arctic.

439 The almost instantaneous turnovers of genetic lineage in *Dicrostonyx torquatus* in Europe and
440 western Russia during the Late Pleistocene has implications for relative dating in the

441 Quaternary. Turnovers, particularly ones involving evolutionary lineages, are considered to be
442 some of the most reliable means of biostratigraphic dating available (Lister, 1992). However,
443 an over-riding concern has been that turnovers are time transgressive and hence cannot be
444 used beyond relatively narrow geographical areas (Lister *et al.*, 2005). The present results
445 showing three rapid turnovers in collared lemmings taking place across more than a 1000 km
446 in Europe and western Russia during the Late Pleistocene may suggest that some of that
447 concern is unnecessary. However, the other implication is that when using turnovers, they
448 may be reliable in the region where the species in question is in its expansive phase but not
449 where it is in its refugium.

450 The ancient sequences from North America suggest that both *D. groenlandicus* and *D.*
451 *richardsoni* probably coexisted in southwestern Canada during the Late Pleistocene (Fig. 1).
452 Identification of *D. groenlandicus* in the Canadian Rocky Mountains is consistent with the
453 most recent morphological identification of collared lemmings from Alberta (Burns, 2004),
454 but recognition of haplotypes indicative of *D. richardsoni* in the Canadian sample reflect
455 greater diversity in Late Pleistocene collared lemming populations than previously
456 recognized. Dates from associated material placed the sequences from January Cave and
457 Eagle Cave between ~33,000 and 40,000 cal. years BP, and >25,000 cal. years BP,
458 respectively (Burns, 1991). Thus, in addition to having a southern distribution as shown by
459 Fulton *et al.* (2013), *D. richardsoni* appears to also have inhabited southwestern Canada
460 during the last glacial period. The glacial origin of *D. groenlandicus* is unresolved but it has
461 previously been proposed that it was restricted to one or more glacial refugia located in the
462 northern part of North America (Ehrich *et al.*, 2000, Fedorov & Goropashnaya, 1999,
463 Fedorov & Stenseth, 2002). Several locations have been proposed for such putative glacial
464 refugia, including ice-free regions in eastern Beringia and the Canadian Arctic Islands or the
465 coastal part of North Greenland, based on paleoecological data (Macpherson, 1965) and

466 modern mitochondrial diversity data (Fedorov & Goropashnaya, 1999, Fedorov & Stenseth,
467 2002). However, in contrast to previous inferences, our findings of *D. groenlandicus* in
468 southwestern Alberta in Canada during the last glaciation suggest that this species may have
469 colonized the high Arctic from a location south of the Laurentide ice sheet following the end
470 of the last glaciation.

471

472 **Conclusions**

473 Reconstruction of the evolutionary history of *Dicrostonyx* across a broad geographic and
474 temporal range represents a unique contribution to our understanding of Pleistocene faunal
475 dynamics. This study demonstrates the potential to reconstruct the evolutionary history of a
476 taxon in detail by analyzing samples from a broad geographical and temporal range. Our
477 analyses unveiled a higher level of genetic diversity across the Late Pleistocene range of the
478 Eurasian collared lemming compared to that observed today. Moreover, an unparalleled
479 pattern of serial genetic replacements through time was revealed across the species' western
480 range during the last 50,000 years. Geographically wide-spread losses of genetic variation and
481 local extinctions during the Late Pleistocene have previously been documented mainly in
482 large-bodied mammals (Barnett *et al.*, 2009, Campos *et al.*, 2010, Lorenzen *et al.*, 2011,
483 Palkopoulou *et al.*, 2013, Stiller *et al.*, 2014). The repeated replacements of collared lemming
484 genetic diversity through time across a large part of its geographical distribution are consistent
485 with a hypothesis that large-scale environmental changes during that time period had a
486 significant impact on the dynamics of cold-adapted small mammals. Additional research is
487 needed to more fully understand the nature of the North American record, and genetic
488 analyses of fossil material from other ecologically similar species will enable us to assess
489 whether such repeated extinctions and replacements were a general pattern among cold-
490 adapted small mammals, which likely would have had cascading effects on the food web of
491 the entire Pleistocene Arctic biota.

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505 **References**

- 506 Abramson NI, Smirnov NG, Tikhonova EP (2004) Morphological studies on collared
507 lemmings (Rodentia, Arvicolidae, *Dicrostonyx*) from Bolshevik Island of the
508 Severnaya Zemlya Archipelago, with notes on evolution and taxonomic position.
509 Russian Journal of Theriology, **3**, 63-70.
- 510 Allen JRM, Brandt U, Brauer A *et al.* (1999) Rapid environmental changes in southern
511 Europe during the last glacial period. *Nature*, **400**, 740-743.
- 512 Baele G, Li WLS, Drummond AJ *et al.* (2013) Accurate Model Selection of Relaxed
513 Molecular Clocks in Bayesian Phylogenetics. *Molecular Biology and Evolution*, **30**,
514 239-243.
- 515 Barnes I, Shapiro B, Lister A *et al.* (2007) Genetic structure and extinction of the woolly
516 mammoth, *Mammuthus primigenius*. *Current Biology*, **17**, 1072.
- 517 Barnett R, Shapiro B, Barnes I *et al.* (2009) Phylogeography of lions (*Panthera leo* ssp.)
518 reveals three distinct taxa and a late Pleistocene reduction in genetic diversity.
519 *Molecular Ecology*, **18**, 1668-1677.
- 520 Blois JL, Mcguire JL, Hadly EA (2010) Small mammal diversity loss in response to late-
521 Pleistocene climatic change. *Nature*, **465**, 771-774.
- 522 Brace S, Palkopoulou E, Dalén L *et al.* (2012) Serial population extinctions in a small
523 mammal indicate Late Pleistocene ecosystem instability. *Proceedings of the National*
524 *Academy of Sciences*, **109**, 20532-20536.
- 525 Brock F, Higham T, Ditchfield P *et al.* (2010) Current Pretreatment Methods for AMS
526 Radiocarbon Dating at the Oxford Radiocarbon Accelerator Unit (ORAU)
527 *Radiocarbon* **52**(1): 103-112 (2010).
- 528 Brown JH (1984) On the Relationship between Abundance and Distribution of Species. *The*
529 *American Naturalist*, **124**, 255-279.

530 Burns JA (1991) Mid-Wisconsinan vertebrates and their environment from January Cave,
531 Alberta, Canada. *Quaternary Research*, **35**, 130-143.

532 Burns JA (2004) Late Pleistocene Lemmings (*Lemmus trimucronatus* and *Dicrostonyx*
533 *groenlandicus*; Muridae: Rodentia) from Alberta, Canada. *Journal of Mammalogy*, **85**,
534 379-383.

535 Campos PF, Willerslev E, Sher A *et al.* (2010) Ancient DNA analyses exclude humans as the
536 driving force behind late Pleistocene musk ox (*Ovibos moschatus*) population
537 dynamics. *Proceedings of the National Academy of Sciences of the United States of*
538 *America*, **107**, 5675-5680.

539 Drummond AJ, Rambaut A, Shapiro B *et al.* (2005) Bayesian Coalescent Inference of Past
540 Population Dynamics from Molecular Sequences. *Molecular Biology and Evolution*,
541 **22**, 1185-1192.

542 Drummond AJ, Suchard MA, Xie D *et al.* (2012) Bayesian Phylogenetics with BEAUti and
543 the BEAST 1.7. *Molecular Biology and Evolution*, **29**, 1969-1973.

544 Ehrich D, Fedorov VB, Stenseth NC *et al.* (2000) Phylogeography and mitochondrial DNA
545 (mtDNA) diversity in North American collared lemmings (*Dicrostonyx*
546 *groenlandicus*). *Molecular Ecology*, **9**, 329-337.

547 Fedorov, Fredga, Jarrell (1999) Mitochondrial DNA variation and the evolutionary history of
548 chromosome races of collared lemmings (*Dicrostonyx*) in the Eurasian Arctic. *Journal*
549 *of Evolutionary Biology*, **12**, 134-145.

550 Fedorov VB (1999) Contrasting mitochondrial DNA diversity estimates in two sympatric
551 genera of Arctic lemmings (*Dicrostonyx*: *Lemmus*) indicate different responses to
552 Quaternary environmental fluctuations. *Proceedings of the Royal Society of London.*
553 *Series B: Biological Sciences*, **266**, 621-626.

554 Fedorov VB, Goropashnaya AV (1999) The Importance of Ice Ages in Diversification of
555 Arctic Collared Lemmings (*Dicrostonyx*): Evidence from the Mitochondrial
556 Cytochrome B Region. *Hereditas*, **130**, 301-307.

557 Fedorov VB, Stenseth NC (2002) Multiple glacial refugia in the North American Arctic:
558 inference from phylogeography of the collared lemming (*Dicrostonyx groenlandicus*).
559 *Proceedings of the Royal Society of London Series B-Biological Sciences*, **269**, 2071-
560 2077.

561 Fulton TL, Norris RW, Graham RW *et al.* (2013) Ancient DNA supports southern survival of
562 Richardson's collared lemming (*Dicrostonyx richardsoni*) during the last glacial
563 maximum. *Molecular Ecology*, **22**, 2540-2548.

564 Geneious version 5.5.7 created by Biomatters. Available from <http://www.geneious.com/>

565 Gill MS, Lemey P, Faria NR *et al.* (2013) Improving Bayesian Population Dynamics
566 Inference: A Coalescent-Based Model for Multiple Loci. *Molecular Biology and*
567 *Evolution*, **30**, 713-724.

568 Golovachov IB, Smirnov NG (2009) The Late Pleistocene and Holocene rodents of the Pre-
569 Urals subarctic. *Quaternary International*, **201**, 37-42.

570 Graham RW (1986) Response of mammalian communities to environmental changes during
571 the late Quaternary. *In: Community Ecology* (eds. Diamond J, Case TJ), Harper and
572 Row, New York., pp. 300–313.

573 Graham RW, Lundelius EL, Graham MA *et al.* (1996) Spatial Response of Mammals to Late
574 Quaternary Environmental Fluctuations. *Science*, **272**, 1601-1606.

575 Hampe A, Petit RJ (2005) Conserving biodiversity under climate change: the rear edge
576 matters. *Ecology Letters*, **8**, 461-467.

577 Hewitt GM (1999) Post-glacial re-colonization of European biota. *Biological Journal of the*
578 *Linnean Society*, **68**, 87-112.

579 Ho SYW, Lanfear R, Phillips MJ *et al.* (2011) Bayesian Estimation of Substitution Rates
580 from Ancient DNA Sequences with Low Information Content. *Systematic Biology*,
581 **60**, 366-375.

582 Hofreiter M, Munzel S, Conard NJ *et al.* (2007) Sudden replacement of cave bear
583 mitochondrial DNA in the late Pleistocene. *Current Biology*, **17**, R122.

584 Kowalski K (1995) Lemmings (Mammalia, Rodentia) as indicators of temperature and
585 humidity in the European Quaternary. *Acta Zoologica Cracoviensia* **38**, 85-64.

586 Leonard JA, Vila C, Fox-Dobbs K *et al.* (2007) Megafaunal Extinctions and the
587 Disappearance of a Specialized Wolf Ecomorph. *Current Biology*, **17**, 1146.

588 Lister AM (1992) Mammalian fossils and quaternary biostratigraphy. *Quaternary Science*
589 *Reviews*, **11**, 329-344.

590 Lister AM, Sher AV, Van Essen H *et al.* (2005) The pattern and process of mammoth
591 evolution in Eurasia. *Quaternary International*, **126-28**, 49-64.

592 Lorenzen ED, Nogues-Bravo D, Orlando L *et al.* (2011) Species-specific responses of Late
593 Quaternary megafauna to climate and humans. *Nature*, **479**, 359-364.

594 Macpherson A (1965) The origin of diversity in mammals of the Canadian arctic tundra.
595 *Systematic Zoology*, **14**, 153-173.

596 Markova AK, Smirnov NG, Kozharinov AV *et al.* (1995) Late Pleistocene distribution and
597 diversity of mammals in northern Eurasia. *Paleontologia I Evolucio*, 5–143.

598 Martin PS (1984) A Prehistoric Revolution. In: *Quaternary Extinctions*. pp. 354-403 (eds
599 Martin PS, Klein RG) Arizona Univ Press, Tuscon.

600 Nadachowski A, Lipecki G, Wojtal P *et al.* (2011) Radiocarbon chronology of woolly
601 mammoth (*Mammuthus primigenius*) from Poland. *Quaternary International*, **245**,
602 186-192.

603 Navascues M, Depaulis F, Emerson BC (2010) Combining contemporary and ancient DNA in
604 population genetic and phylogeographical studies. *Molecular Ecology Resources*, **10**,
605 760-772.

606 Nylander JaA (2004) MrModeltest v2. Program distributed by the author. Evolutionary
607 Biology Centre, Uppsala University.

608 Palkopoulou E, Dalén L, Lister AM *et al.* (2013) Holarctic genetic structure and range
609 dynamics in the woolly mammoth. *Proceedings of the Royal Society B: Biological*
610 *Sciences*, **280**: 20131910.

611 Parks DH, Porter M, Churcher S *et al.* (2009) GenGIS: A geospatial information system for
612 genomic data. *Genome Research*.

613 Prost S, Guralnick RP, Waltari E *et al.* (2013) Losing ground: past history and future fate of
614 Arctic small mammals in a changing climate. *Global Change Biology*, **19**, 1854-1864.

615 Prost S, Smirnov N, Fedorov VB *et al.* (2010) Influence of Climate Warming on Arctic
616 Mammals? New Insights from Ancient DNA Studies of the Collared Lemming
617 *Dicrostonyx torquatus*. *PLoS ONE*, **5**, e10447.

618 Rambaut A, Suchard M, Xie D *et al.* (2014) Tracer v1.6. Available from
619 <http://beast.bio.ed.ac.uk/Tracer>.

620 Ramsey CB (2009) Bayesian Analysis of Radiocarbon Dates. *Radiocarbon* **51**(1):337–360.

621 Reimer PJ, Bard E, Bayliss A *et al.* (2013) IntCal13 and Marine13 Radiocarbon Age
622 Calibration Curves 0–50,000 Years cal BP. *Radiocarbon* **55**(4): 1869–1887.

623 Shapiro B, Drummond AJ, Rambaut A *et al.* (2004) Rise and fall of the Beringian steppe
624 bison. *Science*, **306**, 1561-1565.

625 Sher AV (1991) Problems of the last interglacial in Arctic Siberia. *Quaternary International*,
626 **10–12**, 215-222.

627 Smirnov NG, Fedorov VB (2003) Holarctic collared lemmings: traces of their spread as
628 related to the history of the Arctic biota. *Russian Journal of Ecology*, **34**, 332-338.

629 Stewart JR (2009) The evolutionary consequence of the individualistic response to climate
630 change. *Journal of Evolutionary Biology*, **22**, 2363–2375.

631 Stewart JR, Lister AM, Barnes I *et al.* (2010) Refugia revisited: individualistic responses of
632 species in space and time. *Proceedings of the Royal Society B: Biological Sciences*,
633 **277**, 661-671.

634 Stewart JR, Van Kolfschoten M, Markova AK *et al.* (2003) The Mammalian Faunas of
635 Europe during Oxygen Isotope Stage Three. *In: Neanderthals and Modern Humans in*
636 *the European Landscape during the Last Glaciation, 60,000 to 20,000 years ago:*
637 *Archaeological Results of the Stage 3 Project.* (eds Van Andel TH, Davies SW) pp.
638 103–129. Cambridge, McDonald Institute Monograph Series.

639 Stiller M, Baryshnikov G, Bocherens H *et al.* (2010) Withering Away-25,000 Years of
640 Genetic Decline Preceded Cave Bear Extinction. *Molecular Biology and Evolution*,
641 **27**, 975-978.

642 Stiller M, Molak M, Prost S *et al.* (2014) Mitochondrial DNA diversity and evolution of the
643 Pleistocene cave bear complex. *Quaternary International*, **339–340**, 224-231.

644 Stuart AJ, Kosintsev PA, Higham TFG *et al.* (2004) Pleistocene to Holocene extinction
645 dynamics in giant deer and woolly mammoth. *Nature*, **431**, 684-689.

646 Svensson A, Andersen KK, Bigler M *et al.* (2008) A 60 000 year Greenland stratigraphic ice
647 core chronology. *Climate Of The Past*, **4**, 47-57.

648 Ukkonen P, Aaris-Sørensen K, Arppe L *et al.* (2011) Woolly mammoth (*Mammuthus*
649 *primigenius* Blum.) and its environment in northern Europe during the last glaciation.
650 *Quaternary Science Reviews*, **30**, 693-712.

651 **Supporting Information captions**

652 Supporting Materials and Methods

653 Supporting Table S1. List of sampling localities

654 Supporting Table S2. Details of collared lemming specimens analysed in this study.

655 Supporting Table S3. Details of previously published DNA collared lemming sequences.

656 Supporting Table S4. Primer pairs designed in this study for amplification of the *cyt b* gene.

657 Supporting Figure S1. Date randomization test.

658 Supporting Figure S2. Dated Bayesian phylogeny of mtDNA (*cyt b*) sequences.

659 Supporting Figure S3. Bayesian SkyGrid plot.

660 Supporting Figure S4. Paleoenvironmental records and temporal distribution of the distinct

661 mitochondrial lineages.

662 DNA sequences: Genbank accessions XX-XX

663 **Figure captions**

664 **Figure 1.** Bayesian phylogeny of mtDNA (cyt *b*) haplotypes of *Dicrostonyx* spp. analyzed in
665 this study and map showing their geographic origin. Crosses indicate sites from where
666 modern mtDNA sequences were previously published. Bayesian posterior probabilities of
667 major nodes above 0.8 are shown. Major mitochondrial lineages are indicated by differently
668 coloured branches and vertical dotted lines at the right of the phylogenetic tree. Haplotype
669 numbers refer to Supporting Table S2. Haplotypes mentioned in the text are coloured in red.
670 Symbols indicate the geographic region and age of each haplotype (modern, Late Holocene,
671 Late Pleistocene).

672 **Figure 2.** Bayesian phylogeny of mtDNA (cyt *b*) sequences within *Dicrostonyx torquatus*
673 specimens with finite-radiocarbon dates. Bayesian posterior probabilities of major nodes
674 above 0.8 are shown. The name and median calibrated date of each sequence are given as tip-
675 labels and the temporal range of each mtDNA lineage is shown next to the vertical dotted
676 lines indicating the lineages. Symbols indicate the geographic origin and age of each
677 sequence. The histogram at the bottom shows the number of radiocarbon dated specimens in
678 bins of ~2,000 years. Each bin is coloured according to the proportion of sequences belonging
679 to each of the mtDNA lineages that fall into that bin. The oldest bin '> 46' includes sequences
680 with infinite radiocarbon dates (not included in the phylogeny). The timescale on the x-axis is
681 in thousands of calendar years before present (ky BP).

682 **Figure 3.** Spatial distribution of *Dicrostonyx torquatus* specimens with radiocarbon dates or
683 prior information on their age. Symbol size indicates the number of specimens (below 5,
684 between 5 and 10, and above 10) from a particular site according to the box on the top time-
685 slice. Colours indicate the mitochondrial lineage each specimen belongs to. The two white
686 circles on the top time-slice represent the specimen with the highly divergent sequence from

687 Batagay and the specimen from Bison's site. Time is given in thousands of calendar years
688 before present (ky BP).



- Modern**
- ▲ western Russia
 - ◆ central Russia
 - eastern Russia
 - ▼ North America

- Late Holocene**
- △ western Russia
 - ▣ eastern Russia

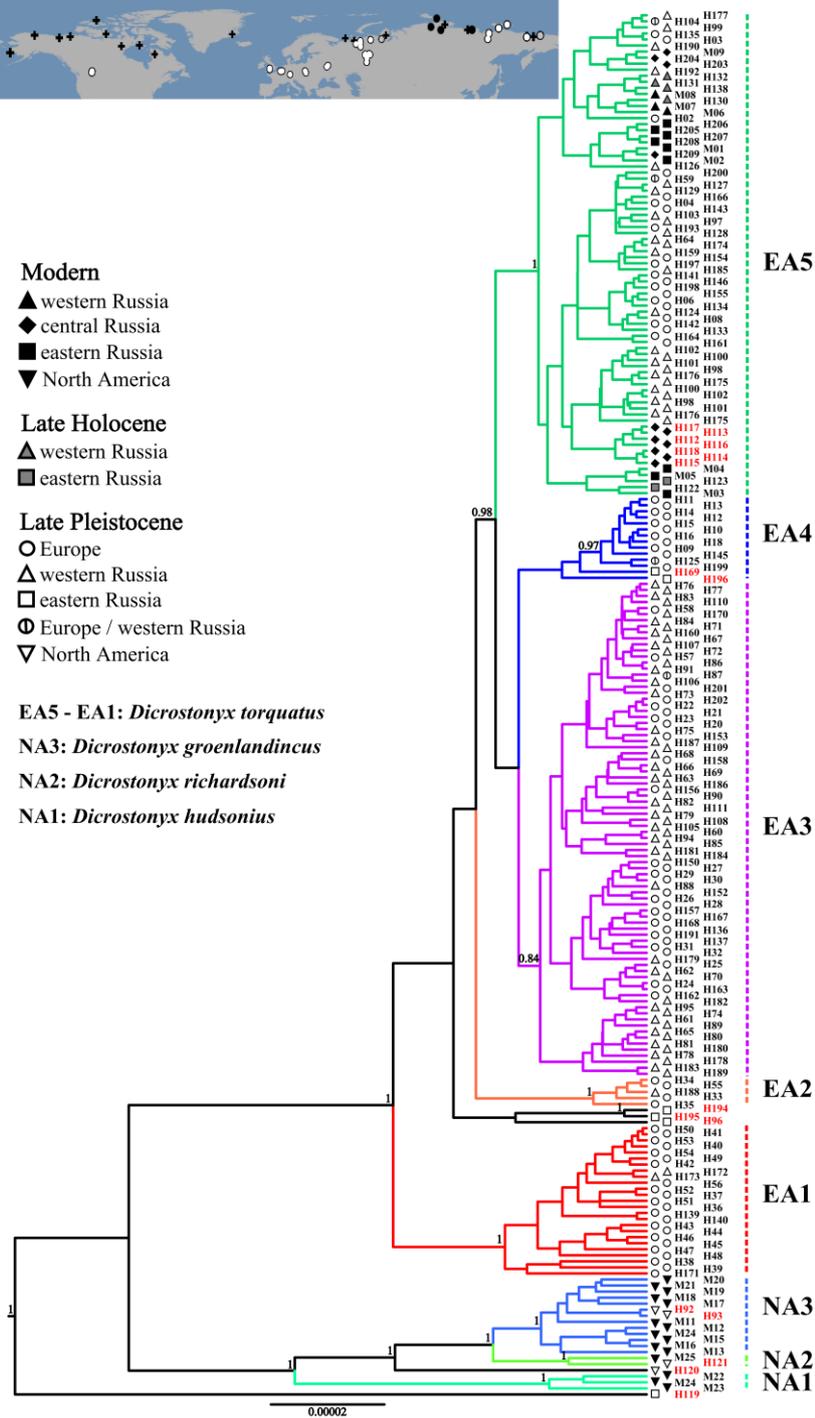
- Late Pleistocene**
- Europe
 - △ western Russia
 - eastern Russia
 - ⊕ Europe / western Russia
 - ▽ North America

EA5 - EA1: *Dicrostonyx torquatus*

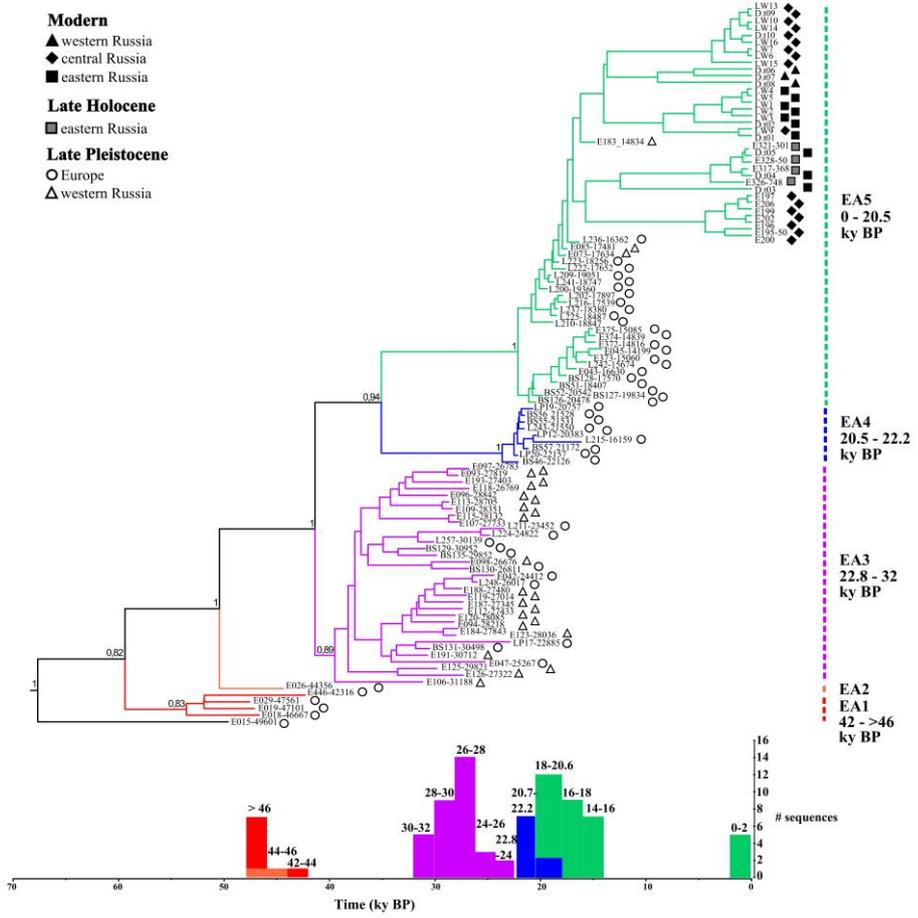
NA3: *Dicrostonyx groenlandicus*

NA2: *Dicrostonyx richardsoni*

NA1: *Dicrostonyx hudsonius*

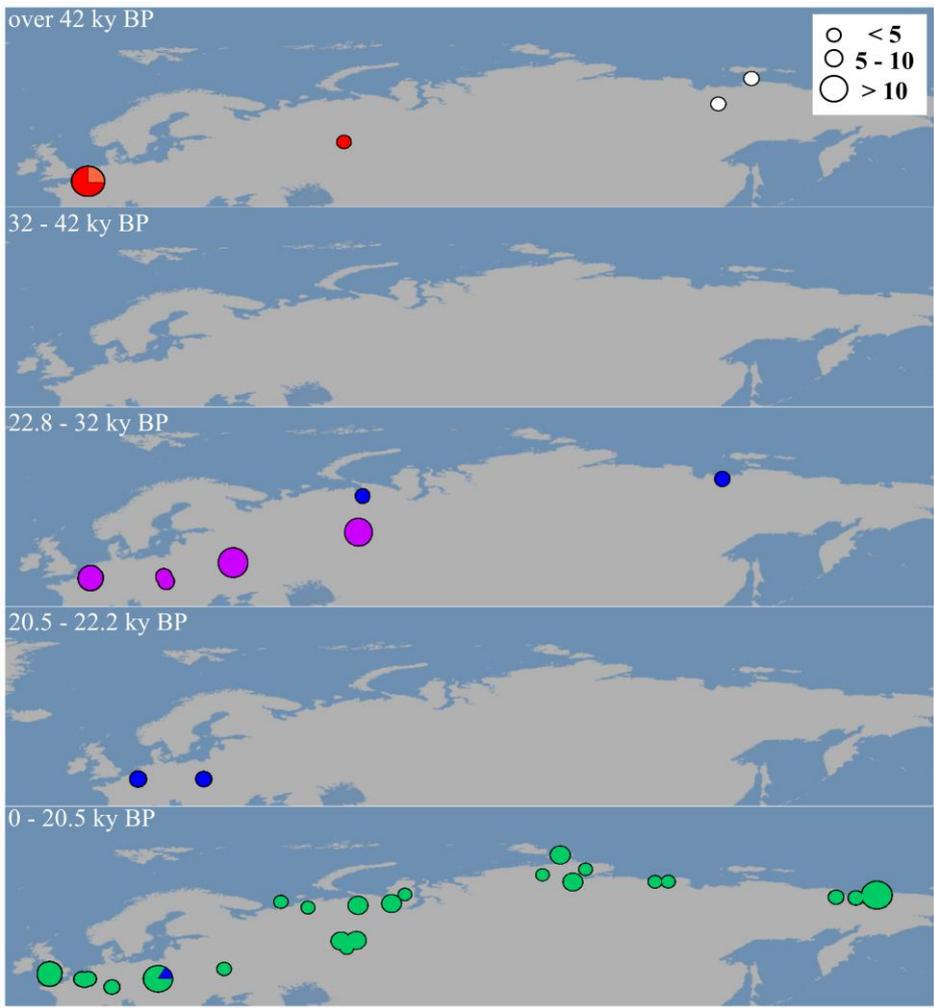


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